Systematic Pore Hydrophobicization to Enhance the Efficiency of an Amine-Based MOF Catalyst – Supporting Information

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MATERIALS

All chemicals were used as received from commercial sources unless otherwise noted. Isopropyl isocyanate (98%), tert butyl isocyanate (97%) and tetradecyl isocyanate (97%) were purchased from Sigma Aldrich. Hexyl isocyanate (99%) and 2-aminoterephthalic acid (BDC-NH₂) (99%) were purchased from Thermo Scientific. Meso- α , β -di(4-pyridyl) glycol (>98%) (DPG) and malononitrile (97%) were purchased from TCI. Sulfuric acid- d_2 (D₂SO₄, 98 wt.% in D₂O, 99.5+% atom D) was purchased from Acros Organics. *N*,*N*-dimethylformamide (DMF), toluene, acetonitrile and benzaldehyde were purchased from Fisher Scientific (ACS certified). Zinc nitrate hexahydrate from Strem Chemicals and 1,4-Diazabicyclo[2.2.2]octane (DABCO) (95%) was purchased from Cambridge Isotopes Laboratories.

SYNTHETIC PROCEDURES

KSU-1 was synthesized according to the literature procedure.²⁶

General procedure for the reaction of MOF materials with isocyanates.

In a typical experiment, ~ 10mg of KSU-1 (0.012mmol) (–OH (0.025mmol) and –NH₂ (0.025mmol)) as-synthesized in DMF was transferred to a 1-dram vial. Then, 2mL of a 0.2 M solution of the isocyanate (0.4 mmol) in acetonitrile were added to the vial. The reaction was left at 80 °C with continuous mixing on a Corning LSE Low Speed Orbital Shaker. Samples of the reacted MOF were taken at 3h, washed in acetonitrile 3 times filtered, and digested for analysis.

General procedure for catalysis

MOF material (0.0075mmol) was introduced into a 0.10 mL glass vial. Benzaldehyde (0.5 mmol; 50.8 uL) and malononitrile (0.55 mmol; 36.5 mg) were then added and the vial was sealed. For reactions in solvent, MOF material (0.0075mmol) was introduced into a 0.10 mL glass vial. Benzaldehyde (0.0625 mmol; 6.35 uL) and malononitrile (0.068 mmol; 4.5 mg) were then added and the vial was sealed 250 μ L of toluene were added, and for those with internal standard, 14.2 μ L (0.08 mmol) of dodecane were also added. The resultant suspension was allowed to react at 50 °C for 30 min. After quenching by cooling to room temperature, the product dissolved with CHCl₃ to separate the solid catalyst. The supernatant was analyzed by ¹H-NMR and the yield determined by integration of signals using disappearance of the starting material to appearance of product.

HIGH-RESOLUTION MASS SPECTROMETRY (HRMS)

MOF samples (~5mg) were placed in a solution of DABCO (10 mg) and 0.25ml of DMSO in a 2-dram vial. The vial was sonicated for ~ 1 min then heated at 80 °C overnight. A small amount of residue was filtered out and the resulting solution was analyzed using a Xevo G2-XS QTof quadrupole time-of-flight mass spectrometer coupled with an ACQUITY M-class UPLC and a NanoLockSpray dual electrospray ion source. Mass spectra were acquired in "eXtreme Resolution" mode.



Figure S1: HRMS of **KSU-1** after reacting with isopropyl isocyanate to form **KSU-1**_{*i*Pr}. Left: the negative mode has no indication of the BDC urea product (m/z = 265.0267), though the [BDC-NH₂-H⁺] starting material peak could also be a fragmentation product. Right: the positive mode has m/z peaks corresponding to [DPG_{dicarbamate}+H⁺] (m/z = 284.1394) and its various fragmentation products.



Figure S2: HRMS of **KSU-1** after reacting with *tert*-butyl isocyanate to form **KSU-1**_{*t*Bu}. Left: the negative mode has no indication of the BDC urea product (m/z = 279.0986), though the [BDC-NH₂-H⁺] starting material peak could also be a fragmentation product (note the presence of an unidentified mass at m/z = 339.1964 that appears to lose successive numbers of $-CH_2$ groups). Right: the positive mode has m/z peaks corresponding to [DPG_{dicarbamate}+H⁺] (m/z = 415.2322) and its various fragmentation products.



Figure S3: HRMS of KSU-1 after reacting with *n*-hexyl isocyanate to form **KSU-1**_{*n*Hex}. Left: the negative mode indicates the presence of the BDC urea product (m/z = 307.1268), and the [BDC-NH₂-H⁺] starting material peak could also be a fragmentation product. Right: the positive mode has m/z peaks corresponding to [DPG_{dicarbamate}+H⁺] (m/z = 471.2960) and its various fragmentation products.



Figure S4:HRMS of KSU-1 after reacting with tetradecyl isocyanate to form **KSU-1**_{C14}. Left: the negative mode indicates the presence of the BDC urea product (m/z = 419.2523), and the [BDC-NH₂-H⁺] starting material peak could also be a fragmentation product. Right: the positive mode has m/z peaks corresponding to [DPG_{dicarbamate}-CH₃⁻] (m/z = 679.5136).

POWDER X-RAY DIFFRACTION (PXRD)

Powder diffraction was recorded on a Bruker AXS D8 Advance Phaser diffractometer (Bruker AXS, Karlsruhe, Germany) with Cu K α radiation (λ = 1.5418 Å) over a range of 4° < 2 θ < 40° in 0.02° steps with a 0.5 s counting time per step. Samples were collected from the bottom of the reaction vial as a thick suspension in DMF and spread on a Si-Einkristalle plate immediately before PXRD measurements.



Figure S5. The PXRD patterns of the simulated **KSU-1**, the experimental **KSU-1**, and the **KSU-1** after reacting with the alkyl isocyanates **KSU-1**_{*i*Pr} and **KSU-1**_{*t*Bu}, **KSU-1**_{*n*Hex}, and **KSU-1**_{*n*C14} respectively.

THERMOGRAVIMETRIC ANALYSIS (TGA)

Thermogravimetric analysis was performed on a TGA 8000 (PerkinElmer Inc., Waltham, MA, USA) interfaced with a PC using Pyris software. Samples were heated at a rate of 10 °C/min under a nitrogen atmosphere. All samples were extensively solvent exchanged with fresh toluene prior to analysis.



Figure S6. TGA data for as-synthesized KSU-1, KSU-1_{*i*Pr} and KSU-1_{*i*Bu}, KSU-1_{*n*Hex}, and KSU-1_{*n*C14} exchanged with toluene.

PROTON NUCLEAR MAGNETIC RESONANCE (1H-NMR)

Spectra were recorded on a Bruker Avance NEO spectrometer (400 MHz for 1H, Bruker BioSpin, Billerica, MA, USA). NMR chemical shifts are reported in ppm against a residual solvent resonance as the internal standard ($\delta(d_6$ -DMSO) = 2.5 ppm). In a typical analysis, MOF materials stored in DMF were solvent exchanged with CHCl₃, isolated by vacuum filtration, and then evacuated in a vacuum oven at 80 °C overnight. Evacuated MOF samples (5-6 mg) were transferred into an NMR tube and d_6 -DMSO (0.55 mL) was added. Subsequently, D₂SO₄ (0.09 mL, 98% w/w in D₂O) was also added. The tubes were capped and sonicated until all the solid was dissolved (~ 1 min).

Dodecane calibration curve

Benzaldehyde (0.5 mmol; 50.8 uL) and dodecane (0.5 mmol; 113.6 uL) were then added into two separate half vial followed by 2.00 mL of toluene in each vial to make a 0.50M solution. These solutions were then diluted to make 0.250 M, 0.200 M, 0.150 M, 0.100 M and 0.0500 M solutions. Aliquots for each solution were analyzed by ¹H-NMR and the ratio of benzaldehyde and dodecane in each standard was determined by integration of dodecane signal at 0.89ppm vs benzaldehyde signal at 10.03ppm.



Figure S7: Calibration curve of the benzylidenemalononitrile proton with dodecane internal standard.

Entry	Catalyst	% Conversion					
		30 min. (I.S.)	30 min. (BA)	3 h (I.S.)	3 h (BA)	6 h (I.S.)	6 h (BA)
1	no catalyst	0	0	0	0	0	0
2	KSU-1	37	37	75	77	90	97
3	KSU-1 _{iPr}	37	37	87	90	96	97
4	KSU-1 _{tBu}	42	44	90	90	96	100
5	KSU-1 _{nHex}	54	57	89	90	96	97
6	KSU-1 _{C14}	58	64	92	90	97	100

Table S1: Comparison of conversions obtained comparing the integral of the benzylidenemalononitrile proton to that of the dodecane internal standard (I.S.) vs comparing to the benzaldehyde (BA) proton.^a

^a0.0625 mmol benzaldehyde, 0.068 mmol malononitrile, 0.083 mmol dodecane, 250 μL toluene,12 mol% catalyst, 50 °C.